

Divergence in morphology, but not habitat use, despite low genetic differentiation among insular populations of the lizard *Anolis lemurinus* in Honduras

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Abstract: Studies of recently isolated populations are useful because observed differences can often be attributed to current environmental variation. Two populations of the lizard *Anolis lemurinus* have been isolated on the islands of Cayo Menor and Cayo Mayor in the Cayos Cochinos Archipelago of Honduras for less than 15 000 y. We measured 12 morphometric and 10 habitat-use variables on 220 lizards across these islands in 2 y, 2008 and 2009. The goals of our study were (1) to explore patterns of sexual dimorphism, and (2) to test the hypothesis that differences in environment among islands may have driven divergence in morphology and habitat use despite genetic homogeneity among populations. Although we found no differences among sexes in habitat use, males had narrower pelvic girdles and longer toe pads on both islands. Between islands, males differed in morphology, but neither males nor females differed in habitat use. Our data suggest that either recent selection has operated differentially on males despite low genetic differentiation, or that they display phenotypic plasticity in response to environmental variation. We suggest that patterns may be driven by variation in intrapopulation density or differences in predator diversity among islands.

Key Words: *Anolis*, Bay Islands, Cayos Cochinos, competition, dewlap, island biogeography, *Norops*, phenotypic plasticity, predation, selection

INTRODUCTION

Research on island organisms has contributed much to our understanding of the selective pressures that shape phenotypic diversity (Losos & Ricklefs 2009, Schluter 2000, Wallace 1902, Whittaker & Fernández-Palacios 2007). Modern comparative studies emphasize the importance of using phylogenetically distinct taxonomic units in an effort to eliminate pseudoreplication (Harvey & Pagel 1991, Hurlbert 1984). Thus, the majority of comparative studies of insular biota have concentrated on endemic species and multi-species radiations (Roughgarden 1995, Schluter 2000), which have often been isolated for hundreds of thousands,

or even millions, of years. While studies that consider phylogenetically independent taxa are useful, they have an often overlooked drawback: the determination of selective mechanisms that have shaped phenotypic diversity in populations over long periods of time (geological scales) is difficult because the role that past ecological conditions play in current morphological or genetic divergence is unknown (Huey & Bennett 1987). By examining multiple populations of one species that have been recently isolated on islands which vary in their ecological conditions, we may reasonably assume that observed differences between these populations are related to current conditions (Garland *et al.* 1991, Grant & Grant 2002, 2003, Schluter 2000, Whittaker & Fernández-Palacios 2007).

In this study, we focused on insular populations of the lizard *Anolis lemurinus*, and build upon a single,

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previous study examining the phylogeographic history of this species in the Bay Islands and Cayos Cochinos of Honduras. Klutsch *et al.* (2007) supported a 'stepping stone' model of archipelago colonization whereby *A. lemurinus* independently colonized the distant islands of Roatan and Utila after first arriving in the Cayos Cochinos from the mainland. Their observations, as well as those of other authors (Kohler 2003, McCranie *et al.* 2005) suggest that the population on Utila is somewhat divergent in general ecology and morphology from those in the Cayos Cochinos and on mainland Honduras, and probably represents a good species. *Anolis lemurinus* in the Bay Islands thus appear to be in the midst of a radiation event. Klutsch *et al.* (2007) also found, however, that populations in the Cayos Cochinos were genetically and morphologically indistinguishable from one another and from mainland populations, suggesting that they have undergone limited evolutionary differentiation or that gene flow between them is ongoing. Nevertheless, these authors did not consider potential differences in habitat use, and only examined coarse aspects of morphology.

This system offered an opportunity to examine ecological and morphological divergence among populations exposed to differing environmental conditions after a relatively recent colonization event. Due to noticeable differences in the abiotic and biotic environments among islands in the Cayos Cochinos, we hypothesized that *A. lemurinus* populations would differ in aspects of morphology and habitat use not considered by Klutsch and colleagues, and that these differences would provide evidence of recent selection or phenotypic plasticity despite genetic homogeneity among populations.

STUDY SITE

Anolis lemurinus occurs on mainland Central America and on the Bay Islands of Honduras (Kohler 2003, Wilson & Hahn 1973). The Bay Islands were formed after the coastal plain of northern Honduras was inundated at the end of the Wisconsin glacial period approximately 10 000 y ago (Bermingham *et al.* 1998). Soon after isolation, mainland *A. lemurinus* colonized a geographically more proximate subset of the Bay Islands known as the Cayos Cochinos (Klutsch *et al.* 2007; Figure 1). *Anolis lemurinus* occurs on the two largest islands of the Cayos Cochinos, Cayo Menor (area = 0.64 km²) and Cayo Mayor (area = 1.67 km²; Bermingham *et al.* 1998, McCranie *et al.* 2005). Cayo Menor and Cayo Mayor sit 18 km from mainland Honduras and are separated by approximately 2.5 km (McCranie *et al.* 2005).

Although Cayo Menor and Cayo Mayor are geographically proximate to one another, they differ noticeably in their biotic environments (Bermingham *et al.* 1998). For example, a species of palm in the

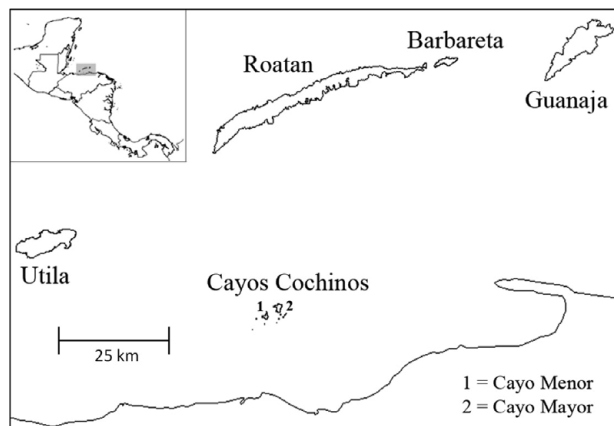


Figure 1. The Bay Islands and Cayos Cochinos of Honduras. Adapted from Green (2010).

genus *Attalea* is particularly abundant on Cayo Mayor, and has noticeably altered understorey and midstorey structure on that island, but does not occur on Cayo Menor (Bermingham *et al.* 1998). Additionally, several species of mammalian and reptilian predators (*Rattus rattus*, *Dasyprocta punctata*, *Leptophis mexicanus*) occur on Cayo Mayor, but are absent from Cayo Menor (Bermingham *et al.* 1998, McCranie *et al.* 2005). Lastly, Cayo Mayor has a permanent source of fresh water, which may increase the carrying capacity or decrease water stress for anoles on that island.

Despite apparent differences in the biotic and abiotic environments of Cayo Menor and Cayo Mayor, very little is known of the morphological or ecological variation of anole populations on these islands. *Anolis lemurinus* is primarily a mainland species (nested within a mainland-derived lineage; Guyer & Savage 1986). As far as we know, they represent the only example of a mainland anole that has successfully colonized an island system in recent geological history (Nicholson *et al.* 2005). Thus, one purpose of our study was simply to document aspects of *A. lemurinus* morphology and habitat use in this archipelago.

METHODS

Sampling procedure

Anolis lemurinus populations on both islands were sampled during the dry season between June and August during 2 y (2008 and 2009). Due to the small size of both Cayo Menor and Cayo Mayor, all areas on both islands with suitable *A. lemurinus* habitat were searched with equal effort. Lizards were captured by hand or by noosing (Blomberg & Shine 2006).

On Cayo Menor all lizards were processed in the laboratory and returned to their respective capture

Table 1. A summary of the morphology of *Anolis lemurinus* on the islands of Cayo Menor and Cayo Mayor in the Cayos Cochinos (mean \pm SE; sample size in parentheses). Snout–vent length, a measure of body length, is abbreviated as SVL.

	Menor		Mayor	
	Male	Female	Male	Female
SVL (mm)	63 \pm 0.6 (76)	64 \pm 0.6 (36)	60 \pm 0.6 (63)	61 \pm 0.4 (45)
Mass (g)	6.0 \pm 0.1 (76)	6.4 \pm 0.2 (36)	5.1 \pm 0.1 (63)	5.4 \pm 0.1 (45)
Tail (mm)	132 \pm 1.9 (43)	123 \pm 2.9 (17)	127 \pm 1.9 (29)	129 \pm 2.2 (25)
Hind-limb (mm)	34 \pm 0.3 (76)	34 \pm 0.3 (36)	32 \pm 0.2 (63)	32 \pm 0.2 (45)
Forelimb (mm)	19 \pm 0.3 (50)	20 \pm 0.3 (25)	19 \pm 0.3 (37)	19 \pm 0.2 (29)
Inter-limb (mm)	23 \pm 0.4 (50)	24 \pm 0.3 (25)	22 \pm 0.4 (37)	22 \pm 0.4 (29)
Jaw length (mm)	17 \pm 0.2 (50)	18 \pm 0.2 (25)	17 \pm 0.2 (37)	17 \pm 0.2 (29)
Jaw width (mm)	10 \pm 0.1 (50)	10 \pm 0.1 (25)	10 \pm 0.1 (37)	10 \pm 0.1 (29)
Pectoral girdle (mm)	8 \pm 0.1 (50)	8 \pm 0.1 (25)	7 \pm 0.1 (37)	7 \pm 0.1 (29)
Pelvic girdle (mm)	5 \pm 0.1 (50)	6 \pm 0.1 (25)	5 \pm 0.1 (37)	5 \pm 0.1 (29)
Toe pad length (mm)	3 \pm 0.1 (50)	3 \pm 0.1 (25)	3 \pm 0.1 (37)	3 \pm 0.1 (29)
Toe pad width (mm)	2 \pm 0.1 (50)	2 \pm 0.1 (25)	2 \pm 0.03 (37)	2 \pm 0.04 (29)
Dewlap length (mm)	17 \pm 0.4 (76)	5 \pm 0.2 (29)	18 \pm 0.4 (63)	5 \pm 0.1 (35)

locations within 48 h. On Cayo Mayor, lizards were processed in the field and released immediately at capture locations. In 2008, to avoid repeat sampling of individuals, each lizard was marked on its ventral surface with a streak of white paint, and no single location was sampled more than twice – once initially, and a second time when a subsample of lizards was returned to a particular location. *Anolis lemurinus* exhibit strong territory affinity and small home-range size (Savage 2002), and these properties were assumed to provide for low re-sampling bias. In 2009, lizards were toe clipped for individual identification. Although we did not permanently mark individuals in 2008, it appears that annual adult mortality in this population is nearly 100% (we did not recapture any individuals during our annual surveys in 2010 and 2011). It is therefore likely that few, if any of the individuals sampled in 2008 were re-sampled in 2009. Additionally, because of large differences in the number of adult versus juvenile lizards sampled on each island, and the potential non-linearity of the relationship between body size and other morphological traits, only adult lizards (snout–vent length \geq 50 mm) were included in this study.

Morphology and habitat use

We measured 12 morphometric variables (Table 1). Mass was measured (to a precision of 0.1 g) using a 10-g-capacity Pesola® spring scale and other morphometric variables were measured (to a precision of 1 mm) with digital calipers. We were interested in potential differences in dewlap size for each population, and in 2008 we measured dewlap length with calipers. We considered ‘dewlap length’ to be the longest distance from the tip of the organ when fully extended to the point where it contacts the chin. We fully extended the dewlap of each

male by clasp the second ceratobranchial cartilages with forceps (Cox *et al.* 2009). However, it was unclear whether dewlap length was a reliable proxy for dewlap area, so in 2009 we calculated dewlap areas for 30 males from Cayo Mayor and 29 males from Cayo Menor. We quantified dewlap area using digital photographs in the image analysis program PhotoJ®, a method that produces highly repeatable results (Vanhooydonck *et al.* 2005a). We hypothesized that dewlap length and area would be tightly correlated so that the former could be used as a proxy for the latter.

At each site where lizards were captured we measured 10 habitat and micro-meteorological variables (Table 2). GPS coordinates, elevation, time of capture, observed behaviour, presence or absence of tail breaks, and sex were also recorded. Sex was determined from an examination of dewlap size and the presence or absence of a hemipenial bulge at the base of the tail. Perch height and distance from the nearest tree whose crown reached the forest canopy were measured with a tape measure. Calipers were used to measure perch diameters ranging from 0 to 15 cm, and diameter tape was used to measure perch diameters greater than 15 cm. Canopy cover was estimated using a concave spherical densiometer. The distance from each observation to the closest open environment was determined with a metric tape measure or estimated visually if that distance was more than 10 m. However, since most individuals were more than 10 m from an open environment, these data lack precision and should be interpreted with caution. An ‘open environment’ was considered to be a canopy gap \geq 5 m². Gaps of this size permit a large amount of solar radiation to reach the forest floor, and likely provide unsuitable microclimates for shade-loving *A. lemurinus* (Savage 2002). ‘Vegetation within 1 m³’ was recorded by visually approximating 1 m³ around the lizard and estimating the per cent volume of that cube containing vegetation. Ambient temperature,

Table 2. A summary of habitat use of *Anolis lemurinus* in the Cayos Cochinos (mean \pm SE; sample size in parentheses). 'Distance to nearest emergent tree' and 'Distance to nearest open environment' are abbreviated 'DNOE' and 'DNOE', respectively.

	Menor		Mayor	
	Male	Female	Male	Female
Perch height (cm)	90 \pm 6.8 (72)	85 \pm 8.8 (34)	105 \pm 6.6 (63)	86 \pm 7.1 (45)
Perch diameter (mm)	69 \pm 8.3 (64)	48 \pm 4.4 (31)	101 \pm 14.7 (59)	76 \pm 12.2 (44)
Canopy cover (%)	97.5 \pm 0.2 (46)	97.7 \pm 0.2 (23)	97.3 \pm 0.2 (37)	98.1 \pm 0.3 (29)
DNET (cm)	35 \pm 6.6 (46)	55 \pm 14.1 (23)	33 \pm 10.0 (37)	40 \pm 11.4 (29)
Proportion vegetation within 1 m ³	0.11 \pm 0.03 (46)	0.09 \pm 0.02 (23)	0.10 \pm 0.03 (37)	0.09 \pm 0.02 (29)
DNOE (m)	28 \pm 3.8 (46)	32 \pm 4.2 (23)	22 \pm 4.4 (37)	18 \pm 3.9 (29)
Ambient temperature (°C)	28.9 \pm 0.2 (72)	28.6 \pm 0.2 (34)	29.0 \pm 0.1 (63)	28.9 \pm 0.2 (45)
Absolute humidity (%)	89.0 \pm 0.9 (46)	90.1 \pm 1.2 (23)	89.0 \pm 0.8 (37)	90.1 \pm 0.9 (29)
Wind speed (m s ⁻¹)	0.1 \pm 0.02 (76)	0.1 \pm 0.03 (36)	0.2 \pm 0.04 (63)	0.3 \pm 0.09 (45)
Solar radiation (μ W cm ⁻²)	24.2 \pm 3.6 (72)	16.5 \pm 1.5 (33)	21.3 \pm 1.9 (63)	20.5 \pm 3.0 (45)

absolute humidity and wind-speed were measured using a Kestrel[®] wind meter. Solar radiation was measured using a Mannix[®] UV light meter.

Statistics

Morphometric and habitat use variables were logarithmically transformed for normality. Sexual dimorphism within and among islands in each morphometric and habitat-use variable was examined using fully-factorial ANCOVAs. The effects of sex, island and all possible interactions were examined for morphology and habitat-use, and non-significant higher-order effects were removed from the final models. For morphometric ANCOVAs, we also included SVL as a covariate to remove the effects of body size. Lastly, a Bonferroni correction was applied to all P-values to correct for inflated Type I error associated with testing multiple hypotheses on the same set of observations (Sokal & Rohlf 1995).

To determine whether morphology and habitat use (as aggregate 'variables' in multivariate space) could be used to distinguish among populations, we performed discriminant analyses on habitat and size-corrected morphometric variables. The effect of size was removed from morphometric variables by generating residual values from regressions of each logarithmically transformed variable on logarithmically transformed SVL. We considered the residuals of mass regressed on SVL to represent an index of body condition. Discriminant functions were based on correlation matrices, and prior probabilities were based on the number of lizards sampled from each island. Coefficients reported are standardized by within-group variances. Due to potential sexual dimorphism and differences in sex ratio among islands, separate discriminant functions were derived for males and females. We derived two discriminant functions for each sex; one for habitat use and one for morphology.

RESULTS

General description of sample

Total sample size consisted of 220 *Anolis lemurinus* (139 males and 81 females). In 2008, we captured 50 males and 25 females on Cayo Menor and 37 males and 29 females on Cayo Mayor. In 2009, we captured 26 males and 11 females on Cayo Menor and 26 males and 16 females on Cayo Mayor. Dewlap length measured with calipers was highly correlated with dewlap area taken from digital photographs of males from both islands in 2009 ($N = 59$, Pearson coefficient = 0.856, $P < 0.001$). We therefore considered dewlap length to be an accurate proxy for dewlap area.

Univariate examination of sexual dimorphism

The maximum SVLs of Cayo Menor males and females were 70 mm and 68 mm, respectively. The maximum SVLs of Cayo Mayor males and females were 67 mm and 66 mm, respectively. Fully factorial ANCOVAs examining sexual dimorphism in *Anolis lemurinus* in the Cayos Cochinos revealed that males had narrower pelvic girdles (significant effect of sex; $F_{1,137} = 23.2$, $P = 0.001$) and longer toe pads (significant effect of sex; $F_{1,137} = 8.78$, $P = 0.048$) on both islands. Additionally, significant interactions between sex and SVL for dewlap length ($F_{1,136} = 8.62$, $P = 0.048$) indicated that this character scaled differently with body length between the sexes. There was also a significant interaction between sex and island for tail length ($F_{1,109} = 12.9$, $P = 0.001$), which indicates that the magnitude of difference between the sexes varied among islands. Sexes did not differ in any of the 10 habitat variables we examined (all $P > 0.160$).

Two additional patterns were revealed through univariate analysis. First, there was an effect of island, but not sex, on mass ($F_{1,216} = 12.1$, $P = 0.012$). Second,

Table 3. Jack-knifed classification matrices for discriminant analyses of males and females using functions derived from either morphological or habitat-use variables, including the percentage of individuals that were classified correctly to their island of origin (last column). An asterisk denotes the function that was able to statistically distinguish among populations.

		Mayor	Menor	% Correct
Male morphology*	Mayor	16	10	55
	Menor	11	36	74
	Total	26	46	67
Female morphology	Mayor	15	10	60
	Menor	11	6	35
	Total	26	16	50
Male habitat-use	Mayor	9	24	27
	Menor	11	28	72
	Total	20	52	51
Female habitat-use	Mayor	17	11	61
	Menor	12	8	40
	Total	29	19	52

body condition was positively correlated with relative hind-limb length (but no other variable) in males (islands pooled; $N = 139$, Pearson coefficient = 0.263, $P = 0.002$), but not females ($P > 0.05$).

Multivariate comparisons among islands

Using discriminant analyses, populations could be distinguished based on male morphology (Wilks' $\Lambda = 0.718$; $F_{12,59} = 1.93$, $P = 0.049$). Relative hind-limb length (coefficient = -0.636), body condition (coefficient = -0.848), and jaw width (coefficient = 0.515) were most heavily weighted in the male-morphology model. Relative dewlap length was moderately weighted in the model (coefficient = 0.286). Classification matrices are displayed for the discriminant functions derived from habitat and morphological variables partitioned among sexes (Table 3). Thus, the discriminant function for male morphology indicates that males on Cayo Menor were in better body condition and had longer hind limbs, narrower jaws and smaller dewlaps at a given SVL than Cayo Mayor males (Figure 2). Females were not distinguishable based on morphology (Wilks' $\lambda = 0.730$, $F_{12,29} = 0.90$, $P = 0.562$), and neither males nor females were distinguishable based on habitat use (Wilks' $\lambda = 0.864$, $F_{10,61} = 0.96$, $P = 0.489$ and Wilks' $\lambda = 0.646$, $F_{10,37} = 2.02$, $P = 0.059$, respectively).

DISCUSSION

Morphological differences between island and mainland populations

The majority of anoles display male-biased sexual size dimorphism (Butler & Losos 2002, Losos 2009, Fitch

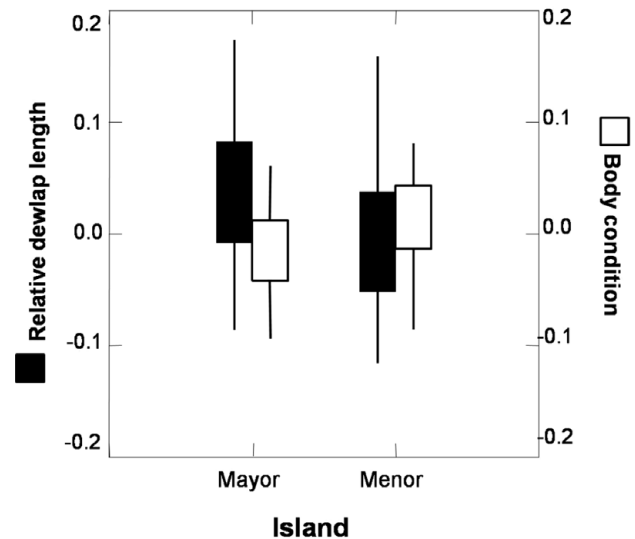


Figure 2. Body condition (indexed as the residuals from a regression of mass on SVL) and relative dewlap length for adult male *Anolis lemurinus* in the Cayos Cochinos.

1976). In contrast, female *A. lemurinus* from mainland Central America are reported to be equal to or greater in size than males (Corn 1981, Savage 2002), and this pattern held for the island populations we studied as well (Table 1). Conversely, Savage (2002) and Guyer & Donnelly (2005) reported little difference in dewlap size among sexes in mainland populations, but male dewlaps were approximately three times larger than that of females in the Cayos Cochinos. Why mainland and island populations differed in this respect is unclear, although sexual dimorphism in dewlap size is often attributed to the role this organ appears to play in territorial defence and courtship behaviour (Jenssen *et al.* 2000, Ord 2008, Vanhooydonck *et al.* 2005a). Additionally, both mainland and island populations of *A. lemurinus* are described as having red dewlaps with black spots (Kohler 2003, McCranie *et al.* 2005, Savage 2002), however, the dewlaps of every individual we observed in the Cayos Cochinos had white spots.

Sexual dimorphism in morphology

Males had longer toe pads and narrower pelvic girdles on both islands, and on Cayo Menor (but not Cayo Mayor) males had longer tails. Broader pelvic girdles in females are consistent with constraints on egg-laying (Castilla & Bauwens 2000). The sex that is more active in the perch matrix is expected to have larger toe pads and longer tails because long tails confer increased balance in arboreal habitat (Gillis *et al.* 2009, Pizzatto *et al.* 2007) and toe pad size is positively correlated with subdigital lamellae number (and therefore clinging

ability; Glossip & Losos 1997, Irschick *et al.* 1996). Increased male activity associated with territory defence and courtship behaviour may provide an explanation for these differences, however, the behavioural data needed to test this hypothesis are unavailable.

Sexual dimorphism in habitat use

Differences in behaviour among male and female anoles often lead to differences in structural or climactic habitat use. In general, this may explain much of the variation in morphology among sexes in *Anolis* lizards (Losos 2009). Nevertheless, male and female *A. lemurinus* in the Cayos Cochinos did not differ in any of the 10 habitat variables we measured.

Differences among islands

Habitat use in multivariate space (discriminant analyses) could not be used to distinguish among populations. This lack of divergence in habitat use is particularly striking given the obvious differences in vegetation structure among islands. Similarity among populations may be a product of niche conservatism (Holt & Barfield 2008, Losos *et al.* 2003, Warren *et al.* 2011), phylogenetic constraint (Harvey & Pagel 1991), or gene flow (Klutsch *et al.* 2007, Lenormand 2002). Unfortunately, because we did not quantify the difference in habitat availability on either island, it is impossible to determine the degree to which lizards are choosing habitat non-randomly.

Despite no discernable difference in habitat use, populations could be discriminated based on male morphology. Cayo Mayor and Cayo Menor males differed in body condition, relative hind-limb length, jaw width and relative dewlap length, as indicated by the coefficients of these variables in the discriminant function derived from male morphological data. Hind-limb length is often positively correlated with perch diameter in anoles as this confers a biomechanical advantage (Calsbeek *et al.* 2007, Irschick & Losos 1999, Langerhans *et al.* 2006). Although males on Cayo Menor had longer hind-limbs than males on Cayo Mayor, we found no difference in perch use among populations. Interestingly, within populations, hind-limb length was positively correlated with body condition, and thus the difference we observed in hind-limb length among islands may have been a result of this correlation.

Differences in male dewlap size and body condition

Surprisingly, males on Cayo Mayor were in worse body condition but had larger dewlaps. We expected lizards that were in better body condition to have more energy to

invest in dewlap development, all else remaining equal. Variation in intraspecific density among islands may explain this pattern. Although abundance data were not collected during the study period, it was apparent from the time required to sample lizards on each island that *A. lemurinus* individuals on Cayo Mayor were much more abundant per unit area than those from Cayo Menor. Males of many lizard species are highly territorial, and at higher densities territorial behaviours such as dewlap extension become increasingly important (Calsbeek & Smith 2007, Lister & Aguayo 1992, Ord 2008). Therefore, males with larger dewlaps might be more successful at defending territories and driving away other males. Additionally, a high population density necessarily results in a reduced per-capita prey base. Thus, density-dependence may be exerting an effect on dewlap size and body condition independently via competitive interactions among males for both territory and prey (Calsbeek 2009, Grether 1996).

The dewlap appears to be an important feature of anole evolution, and several hypotheses (other than population density) have been put forward to explain inter- and intraspecific patterns of dewlap size among anoles (Losos & Chu 1998). Of these, only the 'predator deterrence hypothesis' (Vanhooydonck *et al.* 2009) is congruent with our data. This hypothesis suggests that selection for larger dewlaps should occur when a population is exposed to heightened predation risk because males use their dewlap to advertise escape ability or bite strength to an approaching predator (Vanhooydonck *et al.* 2005b). Of the primary predators of *Anolis* lizards in the Cayos Cochinos, two lizard-specialist snake species (*Leptophis mexicanus* and *Oxybelis aeneus*; Henderson 1982, Savage 2002) occur solely on Cayo Mayor or are at much higher abundance on that island (Bermingham *et al.* 1998, Boback *et al.* 2006, McCranie *et al.* 2005). Moreover, feral cats, dogs and rats (*Rattus rattus*) occur solely on Cayo Mayor and likely add to predation pressure on that island (Bermingham *et al.* 1998, Gasc *et al.* 2010). Thus, it is likely that predation as a whole has been weaker on Cayo Menor where there is a lower diversity of potential predators, supporting the predator deterrence hypothesis as an explanation for dewlap size patterns in the Cayos Cochinos.

Concluding remarks

The factors driving sexual dimorphism among insular *Anolis lemurinus* remain unclear. However, further research into activity budgets and behavioural differences among sexes may reveal potential explanations. Among islands, *A. lemurinus* males differ in several morphological characters, and these differences persist despite no difference in habitat use and a lack of genetic differentiation among populations. As such, we

suggest that differences in anole population density or predation pressure among islands may explain these patterns. Quantitative assessments of anole behaviour and morphology replicated across potentially important environmental gradients within each island are critical for further evaluation of these hypotheses. Our results illustrate the potential for divergent ecological forces to override the homogenizing effect of low genetic differentiation in natural populations.

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LITERATURE CITED

- BERMINGHAM, E., COATES, A., CRUZ, G., EMMONS, L., FOSTER, R. B., LESCHEN, R., SEUTIN, G., THORN, S., WCISLO, W. & WERFEL, B. 1998. Geology and terrestrial flora and fauna of Cayos Cochinos, Honduras. *Revista de Biología Tropical* 46:15–37.
- BLOMBERG, S. & SHINE, R. 2006. Reptiles. Pp. 297–306 in Sutherland, W. J. (ed.). *Ecological census techniques*. Cambridge University Press, Cambridge.
- BOBACK, S. M., MONTGOMERY, C. E., REED, R. N. & GREEN, S. 2006. *Oxybelis aeneus* (Brown Vine Snake). *Herpetological Review* 37: 242.
- BUTLER, M. A. & LOSOS, J. B. 2002. Multivariate sexual dimorphism, sexual selection, and adaptation in Geater Antillean *Anolis* lizards. *Ecological Monographs* 72:541–559.
- CALSBECK, R. 2009. Experimental evidence that competition and habitat use shape the individual fitness surface. *Journal of Evolutionary Biology* 22:97–108.
- CALSBECK, R. & SMITH, T. B. 2007. Probing the adaptive landscape using experimental islands: density-dependent natural selection on lizard body size. *Evolution* 61:1052–1061.
- CALSBECK, R., SMITH, T. B. & BARDELEBEN, C. 2007. Intraspecific variation in *Anolis sagrei* mirrors the adaptive radiation of Greater Antillean anoles. *Biological Journal of the Linnean Society* 90:189–199.
- CASTILLA, A. M. & BAUWENS, D. 2000. Reproductive characteristics of the island lacertid lizard *Podarcis lilfordi*. *Journal of Herpetology* 34:390–396.
- CORN, M. J. 1981. *Ecological separation of Anolis lizards in a Costa Rican rainforest*. Ph.D. thesis, University of Florida, Gainesville.
- COX, R. B., STENQUIST, D. S., HENNINGSEN, J. P. & CALSBEEK, R. 2009. Manipulating testosterone to assess links between behavior, morphology, and performance in the brown anole *Anolis sagrei*. *Physiological and Biochemical Zoology* 82:686–698.
- FITCH, H. S. 1976. Sexual size differences in the mainland anoles. *Occasional Papers of the Museum of Natural History, the University of Kansas* 50:1–21.
- GARLAND, T., HUEY, R. B. & BENNETT, A. F. 1991. Phylogeny and coadaptation of thermal physiology of lizards: a reanalysis. *Evolution* 45:1969–1975.
- GASC, A., DURYEA, M. C., COX, R. M., KERN, A. & CALSBEEK, R. 2010. Invasive predators deplete genetic diversity of island lizards. *PLOS One* 5:e12061.
- GILLIS, G. B., BONVINI, L. A. & IRSCHICK, D. J. 2009. Losing stability: tail loss and jumping in the arboreal lizard *Anolis carolinensis*. *Journal of Experimental Biology* 212:604–609.
- GLOSSIP, D. & LOSOS, J. B. 1997. Ecological correlates of number of subdigital lamellae in anoles. *Herpetologica* 53:192–199.
- GRANT, B. R. & GRANT, P. R. 2003. What Darwin's finches can teach us about the evolutionary origin and regulation of biodiversity. *Bioscience* 53:965–975.
- GRANT, P. R. & GRANT, B. R. 2002. Unpredictable evolution in a 30-year study of Darwin's Finches. *Science* 296:707–711.
- GREEN, S. E. W. 2010. *Evolutionary biology and conservation of the Hog Island Boa constrictor*. Ph.D. Thesis, University of Kent, UK.
- GREETHER, G. F. 1996. Intrasexual competition alone favors a sexually dimorphic ornament in the rubyspot damselfly *Hataerina americana*. *Evolution* 50:1949–1957.
- GUYER, C. & DONNELLY, M. A. 2005. *Amphibians and reptiles of La Selva, Costa Rica, and the Caribbean slope*. University of California Press, Berkeley. 367 pp.
- GUYER, C. & SAVAGE, J. M. 1986. Cladistic relationships among anoles (Sauria, Iguanidae). *Systematic Zoology* 35:509–531.
- HARVEY, P. H. & PAGEL, M. D. 1991. *The comparative method in evolutionary biology*. Oxford University Press, New York. 248 pp.
- HENDERSON, R. W. 1982. Trophic relationships and foraging strategies of some New World tree snakes (*Leptophis*, *Oxybelis*, *Uromacer*). *Amphibia-Reptilia* 3:71–80.
- HOLT, R. D. & BARFIELD, M. 2008. Habitat selection and niche conservatism. *Israel Journal of Ecology and Evolution* 54:295–309.
- HUEY, R. B. & BENNETT, A. F. 1987. Phylogenetic studies of coadaptation: preferred temperatures versus optimal performance temperatures of lizards. *Evolution* 41:1098–1115.
- HURLBERT, S. H. 1984. Pseudoreplication and the design of ecological field experiments. *Ecological Monographs* 54:187–211.
- IRSCHICK, D. J. & LOSOS, J. B. 1999. Do lizards avoid habitats in which performance is submaximal? The relationship between sprinting

- capabilities and structural habitat use in Caribbean anoles. *American Naturalist* 154:293–305.
- IRSCHICK, D. J., AUSTIN, C. C., PETREN, K., FISHER, R. N., LOSOS, J. B. & ELLERS, O. 1996. A comparative analysis of clinging ability among pad-bearing lizards. *Biological Journal of the Linnean Society* 59:21–35.
- JENSSEN, T. A., ORRELL, K. S. & LOVERN, M. B. 2000. Sexual dimorphisms in aggressive signal structure and use by a polygynous lizard, *Anolis carolinensis*. *Copeia* 1:140–149.
- KLUTSCH, C. F. C., MISOF, B., GROSSE, W. R. & MORITZ, R. F. A. 2007. Genetic and morphometric differentiation among island populations of two *Norops* lizards (Reptilia: Sauria: Polychrotidae) on independently colonized islands of the Islas de Bahía Honduras. *Journal of Biogeography* 34:1124–1135.
- KOHLER, G. 2003. *Reptiles of Central America*. Herpeton, Offenbach. 400 pp.
- LANGERHANS, R. B., KNOUFT, J. H. & LOSOS, J. B. 2006. Shared and unique features of diversification in Greater Antillean *Anolis* ecomorphs. *Evolution* 60:362–369.
- LENORMAND, T. 2002. Gene flow and the limits to natural selection. *Trends in Ecology and Evolution* 17:183–189.
- LISTER, B. C. & AGUAYO, A. G. 1992. Seasonality, predation, and the behavior of a tropical mainland anole. *Journal of Animal Ecology* 61:717–733.
- LOSOS, J. B. 2009. *Lizards in an evolutionary tree: ecology and adaptive radiation of anoles*. University of California Press, Berkeley. 528 pp.
- LOSOS, J. B. & CHU, L. 1998. Examination of factors potentially affecting dewlap size in Caribbean anoles. *Copeia* 2:430–438.
- LOSOS, J. B. & RICKLEFS, R. E. 2009. Adaptation and diversification on islands. *Nature* 457:830–836.
- LOSOS, J. B., LEAL, M., GLOR, R. E., DE QUEIROS, K., HERTZ, P. E., SCETTINO, L. R., LARA, A. C., JACKMAN, T. R. & LARSON, A. 2003. Niche lability in the evolution of a Caribbean lizard community. *Nature* 424:542–545.
- MCCRANIE, J. R., WILSON, L. D. & KOHLER, G. 2005. *Amphibians and reptiles of the Bay Islands and Cayos Cochinos, Honduras*. Bibliomania!, Salt Lake City. 210 pp.
- NICHOLSON, K. E., GLOR, R. E., KOLBE, J. J., LARSON, A., HEDGES, S. B. & LOSOS, J. B. 2005. Mainland colonization by island lizards. *Journal of Biogeography* 32:929–938.
- ORD, T. J. 2008. Dawn and dusk “chorus” in visually communicating Jamaican *Anolis* lizards. *American Naturalist* 172:585–592.
- PIZZATTO, L., ALMEIDA-SANTOS, S. M. & SHINE, R. 2007. Life-history adaptations to arboreality in snakes. *Ecology* 88:359–366.
- ROUGHGARDEN, J. 1995. *Anolis lizards of the Caribbean: ecology, evolution, and plate tectonics*. Oxford University Press, Oxford. 226 pp.
- SAVAGE, J. M. 2002. *The amphibians and reptiles of Costa Rica: a herpetofauna between two continents, between two seas*. University of Chicago Press, Chicago. 954 pp.
- SCHLUTER, D. 2000. *The ecology of adaptive radiation*. Oxford University Press, Oxford. 296 pp.
- SOKAL, R. R. & ROHLF, F. J. 1995. *Biometry* (Third edition). W.H. Freeman and Company, New York. 880 pp.
- VANHOODYDONCK, B., HERREL, A., VAN DAMME, R., MEYERS, J. J. & IRSCHICK, D. J. 2005a. The relationship between dewlap size and performance changes with age and sex in a green anole (*Anolis carolinensis*) lizard population. *Behavioral Ecology and Sociobiology* 59:157–165.
- VANHOODYDONCK, B., HERREL, A. Y., VAN DAMME, R. & IRSCHICK, D. J. 2005b. Does dewlap size predict male biting performance in Jamaican *Anolis* lizards? *Functional Ecology* 19:38–42.
- VANHOODYDONCK, B., HERREL, A., MEYERS, J. J. & IRSCHICK, D. J. 2009. What determines dewlap diversity in *Anolis* lizards? An among-island comparison. *Journal of Evolutionary Biology* 22:293–305.
- WALLACE, A. R. 1902. *Island life*. Macmillan, London. 412 pp.
- WARREN, D. L., GLOR, R. E. & TURELLI, M. 2011. Environmental niche equivalency versus conservatism: quantitative approaches to niche evolution. *Evolution* 62:2868–2883.
- WHITTAKER, R. J. & fernández-Palacios, J. M. 2007. *Island biogeography: ecology, evolution, and conservation*. Oxford University Press, Oxford. 416 pp.
- WILSON, L. D. & HAHN, D. E. 1973. The herpetofauna of the Islas de la Bahía, Honduras. *Bulletin of the Florida State Museum* 2:93–150.